Origin of carcinoma associated fibroblasts

Izhak Haviv,^{1,2} Kornelia Polyak,^{3,4} Wen Qiu,^{2,5} Min Hu^{3,4} and Ian Campbell^{5,6,*}

¹Baker IDI Institute; Prahran, Victoria, Australia; ²Department of Biochemistry; ⁶Department of Pathology; School of Medicine; University of Melbourne; Melbourne, Australia; ³Department of Medical Oncology; Dana-Farber Cancer Institute; ⁴Department of Medicine; Harvard Medical School; Boston, Massachusetts USA; ⁵VBCRC Cancer Genetics Laboratory; Peter MacCallum Cancer Centre; East Melbourne, Victoria, Australia

Abbreviations: CAF, cancer associated fibroblasts

Key words: oncology, cancer associated fibroblasts, micro-environment, cancer etiology

Recent evidence on the genomic integrity of non-malignant cells surrounding carcinoma cells has reinvigorated the discussion about the origin of the altered phenotype exhibited by carcinoma associated fibroblasts. Many hypotheses have been proposed for the origin of these altered cells, including standard connective tissue acute phase and stress response, fibroblast senescence, reciprocal interactions with the cancer cells, fibroblast specific somatic mutations, differentiation precursors and infiltrating mesenchymal stem cells. Here we review the definition of CAF phenotype and the evidence for each of those hypotheses, in the context of our current understanding of cancer etiology.

Attempts have been made to reach the goal of medical art-the prevention and cure of disease-by many different paths. New and reliable opportunities have become practicable as our knowledge of the nature of the different diseases has widened (Nobel prize committee presentation speech of the 1945 award for the discovery of Penicillin). One recent discovery that offers an opportunity for novel treatments of cancer involves the cells that are adjacent to epithelial cancer cells, collectively termed stromal cells, and particularly for this review, carcinoma associated fibroblasts (CAFs).¹⁻⁵ CAFs are different from resident fibroblasts of normal tissue in both molecular constitution as well as their functional impact on the neighboring epithelial cells. In animal models of prostate and breast cancer, nonmalignant oncogene-expressing epithelial cells can become malignant when surrounded by fibroblasts that are either oncogene induced, or derived from a primary carcinoma mass.¹ Stroma encoded genes that may modulate the oncogenic potential of adjacent epithelia have been identified,^{2,3} some of which result from signals received from the tumor and some are driven by somatic events within the fibroblast itself. Excellent reviews have been written on the nature of these cells, and their effects on cancer.⁴⁻⁸ In this review we focus on the hypothetical origins of CAFs. Recent evidence shows that unlike

*Correspondence to: Ian Campbell; VBCRC Cancer Research Laboratory; Peter MacCallum Cancer Centre; Locked Bag 1; A'Beckett St; Melbourne, Victoria 8006 Australia; Tel.: +613.96561803; Fax: +613.96561411; Email: ian.campbelll@ petermac.org

Submitted: 12/17/08; Revised: 12/18/08; Accepted: 12/19/08

Previously published online as a *Cell Cycle* E-publication: http://www.landesbioscience.com/journals/cc/article/7669 cancer cells, where many changes occur via somatic mutations, the changes in cancer neighboring cells do not stem from somatic mutations.^{9,10} We will therefore try to consider the reason for this mechanistic disparity between cancer cells and their neighboring cells. "In describing genetic mechanisms, there is a choice between being inexact and incomprehensible" (Nobel prize committee presentation speech of the 1965 award for the discovery of transcription regulation). In this review we shall try to be as inexact as conscience permits, in order to consider as many hypothetical mechanisms as possible for the emergence of CAFs. This aspect of cancer has long been appreciated (desmoplastic stroma¹¹⁻¹³), and has been broadly associated with a poorly differentiated tumor cell phenotype and worse patient outcome¹⁴⁻¹⁷ both in terms of metastatic potential as well as resistance to treatment. Therefore, the origins of CAFs will be considered in both the patient' body as well as from an historical point of view.

Two Parallel Schools in the Research of Cancer Etiology

As research tools developed to allow us to observe smaller and smaller scales of biological structures, it was gradually realized that deformation is the common denominator of a multitude of events in the course of malignancy. Firstly, at the tissue architecture cellular scale in the way cells align relative to neighbors to form tissues, and migrating cells such as inflammatory cells infiltrate the tissue, in the cellular and nuclear morphology and, as noticed by Bovery,¹⁸ in the molecular scale of genomic integrity. The cause and effect relationship between the molecular genomic level and that of the cellular level fits the paradigm of nature versus nurture conflict of perspectives, and not surprisingly is the focus of substantial debate that perpetuates along the history of cancer research.

Genomic Deformation

While the clinical and practical definition of the cancer is morphological, most cancer researchers believe, in spite of opposition,¹⁹ that cancer is the outcome of DNA-mutational events.²⁰ The DNA-centered view of cancer etiology begins with the discovery that cancer cells carry chromosomal imbalance,¹⁸ that chromosomes carry our heritable material,²¹ that DNA therein is the molecule of heredity,²² and that somatic DNA changes occur in and modulate the behavior of cancer cells.^{23,24} It is hard to strictly apply Koch's postulates to the etiology of cancer; while organisms with inherited

oncogenic mutations often develop into cancer prone individuals, the mere viability of these mutants suggests that cancer is a complex multigenic syndrome. Single oncogene-transformed cells still require additional changes in order to exhibit the phenotype of full blown cancer.^{25,26} Correlating with this, the majority of cells from premalignant human tissues already harbor multiple mutations^{27,28} in that precursor state. Nevertheless, in a broader sense the DNA-centered view of cancer fulfils all Koch's postulates. Mutations found in cancer have been successfully isolated from a diseased organism, 24,29,30 have been shown to cause disease when introduced into a healthy organism,^{31,32} and usually the sustainable presence of these mutations is necessary for maintenance of the disease state.^{33,34} Thus, with some exceptions, re-deriving the mutation from the disease state is possible at any stage of the disease. The phenotype of a cancer cell involves multiple incremental changes³⁵ that accumulate in an escalating manner through extensive in vivo selection^{25,36,37} and evolutionary dynamics.³⁸⁻⁴⁰ Furthermore, the one-gene-onefunction paradigm is systematically substituted with models where multiple gene products regulate a single phenotype, and single hits in any of those independent loci can give rise to the same outcome. Consequently, cancer causing mutations, from an etiological point of view, may be better categorized into pathways. If so, then even the first of Koch's postulates, i.e., that the oncogenic mutations must be found in all individuals suffering from the disease is fulfilled if you collate them into pathway groups. Indeed this is the case for canonical pathways, such as RAS and PI3K, where any given mutation is present in an average of 5% of the patients, and yet those pathways are deregulated in almost all cases.^{26,41} A bigger discrepancy with Koch's postulates is the finding that many premalignant conditions carry such mutations, somehow without the full blown disease.⁴²⁻⁴⁵ Extrinsic control of tumorigenesis and progression, which likely affect the genomic integrity indirectly via the selective pressure that premalignant cells are exposed to could therefore demark the premalignant to malignant transition.^{19,46,47}

Cellular Deformation

The first notion of stromal control of cancer comes from the microscopic observation of Virchow that there is a strong association between morphological features of wounded tissue and inflammation and cancer (elevated levels of infiltrating inflammatory and immune cells).^{48,49} Inflammation is also associated with an elevated risk of cancer.49-52 In addition, by examining more than 900 autopsy records of patients with different primary tumors, Paget documented a non-random pattern of metastasis to visceral organs and was struck by the discrepancy between the relative blood supply and the frequency of metastases in certain organs, such as breast and prostate cancer spread to bones. This lead him to suggest what is now termed the "seed and soil" hypothesis, which assigns a role for both cancer and stromal cells in the establishment of distant metastasis, and consequently organ specific metastatic destiny preference.^{53,54} In 1951, it was shown that skin irritation by carcinogen increased the efficiency of carcinogen-unexposed epithelial cell transformation.⁵⁵ Even in the context of cells expressing an activated oncogene, tumor formation still depended on wounding.^{56,57} This suggested that carcinogenesis is reliant on higher order cell-cell interactions rather than on a simple cell autonomous DNA damage phenomena as suggested by Ames and colleagues.⁵⁸

Integrating the Two Levels of Deformation in Cancer

Normal solid tissue is composed of multiple cell lineages, such as endothelial cells which constitute blood vessel walls, epithelial cells that make up body surfaces and provide glandular functions, and fibroblasts which comprise and direct the maintenance of connective tissue and the extracellular matrix. Between these major cell lineages, there is a dense protein barrier called basement membrane. To ensure that the basement membrane is properly positioned at the interface, heterotypic interactions between those cell lineages induce the mutual deposition of basement membrane components, laminins or collagens, by the epithelial or fibroblast cells, respectively. This barrier not only demarks the location of cells according to lineage commitment, but also plays a critical role in coordinating the functions, life span and behavior of the cells mainly through molecular attachment. As a result of this symbiotic mutual dependence of metazoan cells, the overall function of the organism depends on proper ratios and relative positioning of different cell types in the space of tissues. This remarkably complex tissue structure is achieved by a coordinate choreography, during which cells propagate and acquire identity concomitantly in response to their tissue location (Spemann-Mangold organizer effect as paradigm).⁵⁹⁻⁶¹ Perturbation to this morphological homeostasis, and the basement membrane in particular, either via mechanical injury or enzymatic degradation, induces a response generally termed as wound healing or inflammation. The desired outcome of this homeostatic response is the recovery of proper tissue structure and function. A malignant cell that invades and destroys this barrier violates this vital tissue architecture and morphological equilibrium and elicits a perpetual wound healing response in the cancer microenvironment.⁶² In the context of malignancy, the wound healing response turns the invaded stroma from harmless boundary into an active tumor promoter.

Evolutionary Context for Cancer Promotion by Spontaneous Inflammatory Reaction

"Everywhere in nature we observe adaptations to the finest degree one can think of" (Schroedinger, 1998). It therefore seems counterintuitive that an unaltered healthy stroma would respond to the cancer in a manner that ameliorates the pathology and become an accomplice by spontaneous response, without a change in the stromal cells themselves. Yet, spontaneous cross talk between epithelial and stromal cells induces the expression of genes in both the stroma and cancer cells.⁶³⁻⁷¹ These genes include classical cancer stroma markers that reportedly may promote oncogenic potential of adjacent epithelia.^{2,3,72-74} This is possibly due to the late presentation with carcinoma disease in human life span. Diseases that present in individuals at post reproductive age, are not expected to be selected against in human evolution.⁷⁵ By contrast, inflammation, which is an acute response to infection provides evolutionary advantage at reproductive age. Since recovery of the epithelial cell function is critical to wound healing, inflammation activates proto-oncogenes,⁷⁶ increases genomic instability, via oxidative radicals^{52,77} and protects oncogene-transformed epithelial cells from apoptosis.^{50,51,78} Considering the microevolution of cancer cells via somatic alterations, at the rate of somatic mutations observed in normal cells, it is difficult to envisage how cancer cells get to accumulate their typical havoc genome damage. It therefore follows that events

early in premalignant conditions⁷⁹ lead to an intrinsic increase in mutation rate, as well as loss of mechanisms that monitor genomic integrity and control the appropriate cellular response, such as DNA repair or cell death. Since inflammation accelerates almost all of the milestones of cancer progression, it follows that chronic inflammation is linked with increased risk of a few different cancer types. We consider the CAFs as part of these cancer microenvironment changes that occur in tissue lesions and serve as precursors for malignant disease. Several hypotheses have been presented for the origin of these altered cells, including standard connective tissue acute phase and stress response,^{55,80,81} and fibroblast senescence,⁸²⁻⁸⁵ reciprocal interactions with the cancer cells,^{5,20,86-90} fibroblast specific somatic mutations,⁹¹⁻⁹⁵ differentiation precursors and infiltrating mesenchymal stem cell.^{96,97}

Connective Tissue Acute Phase and Stress Response Model of CAFs

The first investigation of the role of altered stroma in cancer etiology was reported as early as 1951 where transplanted carcinogen (methylachollantrene)-treated skin mesenchymal cells were shown to induce an increased incidence of skin carcinoma. While these early experiments did not have methodologies to exclude the possibility that the cancers arose from contaminating epithelial cells, they nevertheless raised the compelling hypothesis that carcinogens affect tumor stroma, which then plays an initiator role in cancer etiology. More recently, this experiment was reproduced in rats for breast cancer,⁸¹ however, again, the data lacks definitive proof that the malignancy is not a product of direct mutagenesis of epithelial cells contaminating the stromal adaptive transfer. It is also not clear if the cancer promotion was due to carcinogen induced somatic mutations or simply stress-related alterations in gene expression.^{80,98,99}

The Fibroblast Senescence Model of CAFs

One of the prime barriers to oncogenic transformation is the limited license to proliferate in vitro that differentiated somatic cells exhibit, which means that by enlarge, elevated proliferation is unsustainable.¹⁰⁰⁻¹⁰⁵ Cancer usurps this barrier by various intracellular mechanisms, but on the level of cancer microenvironment, senescence may also promote cancer.¹⁰⁶⁻¹⁰⁸ The linear increase in cancer incidence with age fits the model of cumulative somatic mutations for cancer etiology¹⁰⁹⁻¹¹² but other contributions of age are possible. In particular, age is correlated with generalized chronic increase in tissue inflammation. 49,50,85,107,113,114 Since senescent fibroblasts spontaneously express a host of inflammatory cytokines linked with cancer promotion⁸⁵ it has been proposed that normal fibroblasts of aging individuals play an initiating role in cancer etiology through an inherent CAF phenotype.¹¹⁴ The majority of reports for CAFs however record that CAFs promote cancer to a greater extent than fibroblasts derived from distant tissue within the same patient, suggesting that CAFs arise from further biological events. Additionally, expression profiles of senescent fibroblasts partially, but not completely phenocopy the expression signatures of CAFs. Nevertheless, since proliferation accelerates senescence, it is possible that moderate increase in local mitogenic signals to the fibroblasts within cancer microenvironment (see next model), contribute to specific accumulation of senescent fibroblasts within cancer microenvironment beyond the normal tissues of the patient.

The Reciprocal Interactions Model of CAFs

In the cancer epithelial cell-centered model for carcinogenesis, CAFs are merely an inevitable response to the cancer causing mutations and are fundamentally no different from normal fibroblasts (as part of the inflammatory reaction described above).¹¹⁵⁻¹¹⁸ The fibroblast response is hardwired in the genome as part of the cancer's resemblance to a chronic wound, aiming at support of epithelial cell survival and expansion.¹¹⁹⁻¹²⁸ In addition to parsimony, this hypothesis offers clear predictions to scientifically test against corresponding null hypotheses; (1) That co-culture of cancer cells with normal fibroblasts will induce expression of CAF-specific genes in the fibroblasts,⁶³⁻⁷¹ (2) that wounded fibroblasts should promote cancer in a way that is indistinguishable from CAFs,¹²⁹ and (3) that normal fibroblasts can transform into CAFs via co-cultivation with cancer cells in vivo for extended period of time (our unpublished work does not provide evidence for this prediction). Gene expression profiling of tumor-stromal interactions between co-cultured cancer cells and stromal fibroblasts have previously been performed for cancer cells with the corresponding organ-specific fibroblasts.⁶³⁻⁷¹ Many of the genes shown to be activated in these co-cultures are known markers of CAFs in vivo, such as MMP1, MMP3, collagens, TNC, etc. Evidence that this reciprocal interaction promotes cancer includes the anti-cancer effect of Imatinib, on carcinoma animal models. It was shown that inhibition of the PDGFRB in cancer stroma leads to attenuated bFGF signaling, and consequent attenuation of the carcinoma.¹³⁰ However, many of the genes that are elevated in CAFs do not undergo a change of expression in co-cultures. It is possible that some other cells in the tumor microenvironment and missing in the co-cultures, such as tumor associated macrophages, play a key role in regulating the phenotype of CAFs. Alternatively, some other aspect of the CAF-origin is not recapitulated by co-culture experiments.

The Mutational Model of CAFs

The impressive progress in the identification and characterization of tumor-causing mutations in oncogenes or tumor suppressor genes has in a sense indoctrinated our thinking about the underlying molecular basis of alterations in cell behavior.^{26,35,131} Following the success with genome analysis of the cancer cell itself, it was only natural to use the same hypotheses for the exploration of all aspects of cancer, including the cancer microenvironment. Molecular genetic studies in breast cancer have reported somatic mutations in TP53 and PTEN as well as gene copy number alteration at other loci in adjacent stroma^{91,93,132} suggesting that much of the tumor promoting activity of stromal cells may be mutation based. The co-existence of mutations in two (or more) cell lineages, was initially claimed to be the product of sequential mutagenesis, where the oncogenic mutations occurred in the cancer cell first, and then in the consequent host infiltrate, as an inevitable outcome.²⁰ It was also suggested that multiple cell lineages concomitantly incur somatic mutations that favored cancer promoting symbiotic relationships between the cancer cell and the adjacent supporting connective tissue.⁹⁹ However, this concept is still contentious.¹³³ Considering the potential increased mutation rate in inflamed microenvironment, it is possible that the resident stromal cells could accumulate a random set of mutations. On the other hand, the classical working hypotheses on carcinogenesis assumes the mutation rate is affected by overall rate of proliferation,

which would be orders of magnitude lower in the stromal cells compared to the cancer cells. It was nevertheless exciting to see that somatic alterations were consistently observed at a high frequency (>30%) in tumor juxtaposed fibroblasts suggesting that these were the underlying molecular basis for the sustained cancer promoting attributes of CAFs.^{91,93} Unfortunately, technical aspects of this body of work raised serious doubt as to whether these apparently frequent somatic mutations were genuine.¹³⁴ Our own studies have revealed only a single loss of heterozygosity event on chromosome 22 in one CAF population among 35 breast and ovarian cancers. Our data and the consensus of molecular genetic studies of CAFs where technical artifacts were avoided is that somatic alterations in clonal populations of CAFs are at best exceedingly rare.^{9,10}

Hypothetically, the idea of co-evolution of two cell lineages in the body that carry independent somatic mutations is not implausible. An altered fibroblast that somehow entices the neighboring epithelia to secrete a support signal for other fibroblasts would be expected to consequently expand further. The altered fibroblast might propagate more rapidly than distant normal fibroblast counterparts since it is the source of the supporting environment and the effect might be expected to be much localized. However, in such scenario, unaltered fibroblasts immediately adjacent to this altered fibroblast are expected to expand at a similar rate in a manner reminiscent to satellite bacterial colonies that emerge in selective bacterial culture if the ampicillin selection was extended so long that the secreted β -lactamase fully eliminated the ampicillin in the vicinity of a resistant colony. This model predicts that if CAF mutations existed they would generate clonal expansion in the midst of unaltered bystander normal fibroblasts and they would not reach homogeneity of mutant cells. This means that even if a mutant fibroblast that promotes cancer was to benefit from this mutation directly, it might not be detectable with current technologies. If this co-evolution model of CAF mutations is correct, one would still expect to observe evidence for fibroblast proliferation which would be needed to drive the clonal expansion. However, Ki67 and PCNA staining of tumor sections invariably failed to detect proliferating fibroblasts. Similarly, in instances where immunohistochemistry is capable of detecting mutant proteins, such as p53, the signal is invariably centered on the cancer cells. Of course, speckled signal is sometimes observed for the stroma, but not necessarily above background. Overall, any model which incorporates somatic mutations in CAFs is not supported by empirical data and in our view is not a tenable explanation for the CAF phenotype.

Differentiation Intermediates and Mesenchymal Precursors Model of CAFs

The comparison between cancer and wounds⁶² is based on the fact that both tissue regeneration and carcinogenesis involve cell proliferation, survival and migration that are controlled by growth factors and cytokines as well as inflammatory and angiogenic signals. In particular, tissue injury leads to acute recruitment of immune cell infiltrates, which are early markers of basement membrane breakdown, followed by more sustainable fibrosis.^{135,136} Whereas it is possible that this fibrosis results from responses in local fibroblasts, mesenchymal precursors are also known to be recruited to injured tissue,¹³⁷ as well as cancer.¹³⁸ Most compellingly, knee aspirate-derived human mesenchymal stem cells were shown to support not only primary cancer growth,⁹⁷ but most importantly overall breast cancer metastasis.

Unfortunately, these experiments were not performed side-by-side with CAFs and normal fibroblasts from the same individuals. Since the activity of precursor cells is defined by in vitro colony forming capacity, it is hard to predict the expected result in the case that these cells are indeed the source of CAFs in cancer tissue. The simple prediction would be that a large fraction of the fibroblastic cell population in tumors would be progeny of mesenchymal precursor cells that migrated from an external body pool. While some evidence suggest this indeed is the case,138 using BGal+ ROSA 26 bone marrow-derived mesenchymal cells we only observed small numbers of such progeny in tumor cross sections (manuscript in preparation). Precursor cells may convey tissue regeneration by coordinating the proliferation and migration of other cells, and play a critical role in the process, without contributing a large fraction of the ultimate cell numbers. In fact, in the case of tumor vasculature it was observed that endothelial precursor cells only produce a small fraction of progeny cells that are recognized within tumor vasculature, yet their infiltration into tumors is critical for the overall formation of tumor vasculature.¹³⁹ Whilst this is a plausible model, there is no evidence for this model to date. Another source of differentiation intermediates potentially contributing to CAF activity may be from tissue pericytes. These peri-vascular cells share a large number of cell markers with CAFs including PDGFRB, Thy-1 and NG2. In tumors however, these markers are not restricted to the peri-vascular position as they are in normal tissues.¹⁴⁰ Evidence supporting the link between pericytes and CAF-like cancer support comes from analysis of the effect of STI-571 in mouse carcinoma models.¹³⁰ While these models are compelling, the ultimate validation would come from animal model experiments where adaptive transfer of mesenchymal precursor, either from the bone or normal tissue, could be assayed for the capacity to promote cancer growth as compared to original CAFs.

Concluding Remarks

It is important to note that the models described in this review are not mutually exclusive, both across different patients, as well as across different fibroblastic cells in any given tissue since cell marker studies indicate that fibroblasts are quite heterogeneous.¹⁴⁰ The heterogeneity of the tissue fibroblasts is critical, since the biological activity that CAFs convey onto to cancer has not been purified to homogeneity, nor is there an assessment of the specific activity for these assays, i.e., how many of the fibroblasts deliver the functional effect, in the background of irrelevant fibroblast cells. Thus, either of those populations could be responsible for the observed support of cancer, in addition to expressing distinct sets of gene products. This also means that the tumor stromal cells that promote cancer in the animal assays for CAFs may in fact carry a unique set of somatic mutations, which were obscured by the diploid genome of the majority of the stromal fibroblasts, which may be viewed as innocent bystanders. However, this possibility cannot explain the previous publications claiming accumulation of mutations in the bulk carcinoma fibroblasts, nor can this possibility be addressed until the cells that promote cancer are further purified to homogeneity.

Acknowledgements

Dr. Polyak reports receiving consulting fees from Novartis, Pfizer and AVEO Pharmaceuticals, holding stock in AVEO Pharmaceuticals, receiving lecture fees from Biogen Idec, and receiving grant support from Novartis and Biogen Idec.

References

- Olumi AF, Grossfeld GD, Hayward SW, Carroll PR, Tlsty TD, Cunha GR. Carcinomaassociated fibroblasts direct tumor progression of initiated human prostatic epithelium. Cancer Res 1999; 59:5002-11.
- Inaguma Y, Kusakabe M, Mackie EJ, Pearson CA, Chiquet-Ehrismann R, Sakakura T. Epithelial induction of stromal tenascin in the mouse mammary gland: from embryogenesis to carcinogenesis. Dev Biol 1988; 128:245-55.
- Paley PJ, Goff BA, Gown AM, Greer BE, Sage EH. Alterations in SPARC and VEGF immunoreactivity in epithelial ovarian cancer. Gynecol Oncol 2000; 78:336-41.
- Bhowmick NA, Moses HL. Tumor-stroma interactions. Curr Opin Genet Dev 2005; 15:97-101.
- Bierie B, Moses HL. Tumour microenvironment: TGFbeta: the molecular Jekyll and Hyde of cancer. Nat Rev Cancer 2006; 6:506-20.
- Micke P, Ostman A. Tumour-stroma interaction: cancer-associated fibroblasts as novel targets in anti-cancer therapy? Lung Cancer 2004; 45:163-75.
- Mueller MM, Fusenig NE. Friends or foes—bipolar effects of the tumour stroma in cancer. Nat Rev Cancer 2004; 4:839-49.
- Orimo A, Weinberg RA. Stromal fibroblasts in cancer: a novel tumor-promoting cell type. Cell Cycle 2006; 5:1597-601.
- Qiu W, Hu M, Sridhar A, Opeskin K, Fox S, Shipitsin M, et al. No evidence of clonal somatic genetic alterations in cancer-associated fibroblasts from human breast and ovarian carcinomas. Nat Genet 2008; 40:650-5.
- Campbell IG, Qiu W, Polyak K, Haviv I. Breast-cancer stromal cells with TP53 mutations. N Engl J Med 2008; 358:1634-5.
- Dvorak HF, Dickersin GR, Dvorak AM, Manseau EJ, Pyne K. Human breast carcinoma: fibrin deposits and desmoplasia. Inflammatory cell type and distribution. Microvasculature and infarction. J Natl Cancer Inst 1981; 67:335-45.
- 12. Spain DM. The association of terminal bronchiolar carcinoma with chronic interstitial inflammation and fibrosis of the lungs. Am Rev Tuberc 1957; 76:559-66.
- Rutter AG. Idiopathic retroperitoneal fibrosis simulating advanced pelvic carcinoma. Br J Urol 1965; 37:302-6.
- Chang HY, Nuyten DS, Sneddon JB, Hastie T, Tibshirani R, Sorlie T, et al. Robustness, scalability and integration of a wound-response gene expression signature in predicting breast cancer survival. Proc Natl Acad Sci USA 2005.
- Segal E, Friedman N, Koller D, Regev A. A module map showing conditional activity of expression modules in cancer. Nat Genet 2004; 36:1090-8.
- Anscher MS, Peters WP, Reisenbichler H, Petros WP, Jirtle RL. Transforming growth factor beta as a predictor of liver and lung fibrosis after autologous bone marrow transplantation for advanced breast cancer. N Engl J Med 1993; 328:1592-8.
- Antoniades HN, Galanopoulos T, Neville-Golden J, O'Hara CJ. Malignant epithelial cells in primary human lung carcinomas coexpress in vivo platelet-derived growth factor (PDGF) and PDGF receptor mRNAs and their protein products. Proc Natl Acad Sci USA 1992; 89:3942-6.
- 18. Boveri TM. Concerning the origin of malignant tumours. J Cell Sci 1902; 121:1-84.
- Sonnenschein C, Soto AM. Somatic mutation theory of carcinogenesis: why it should be dropped and replaced. Mol Carcinog 2000; 29:205-11.
- 20. Bernards R, Weinberg RA. A progression puzzle. Nature 2002; 418:823.
- 21. Morgan TH. Chromosomes and associative inheritance. Science 1911; 34:636-8.
- Avery OT, MacLeod CM, McCarty M. Studies on the chemical nature of the substance inducing transformation of pneumococcal types: induction of transformation by a Deoxyribonucleic acid fraction isolated from pneumococcus type III. J Exp Mrf 1944; 79:137-58.
- Knudson AG. Mutation and cancer: Statistical study of retinoblastoma. Proc Natl Acad Sci USA 1971; 68:820-3.
- Varmus HE, Quintrell N, Medeiros E, Bishop JM, Nowinski RC, Sarkar NH. Transcription of mouse mammary tumor virus genes in tissues from high and low tumor incidence mouse strains. J Mol Biol 1973; 79:663-79.
- Halachmi E, Witz IP. Differential tumorigenicity of 3T3 cells transformed in vitro with polyoma virus and in vivo selection for high tumorigenicity. Cancer Res 1989; 49:2383-9.
- Sjoblom T, Jones S, Wood LD, Parsons DW, Lin J, Barber TD, et al. The consensus coding sequences of human breast and colorectal cancers. Science 2006; 314:268-74.
- Aubele M, Cummings M, Walsch A, Zitzelsberger H, Nahrig J, Hofler H, et al. Heterogeneous chromosomal aberrations in intraductal breast lesions adjacent to invasive carcinoma. Anal Cell Pathol 2000; 20:17-24.
- Berman H, Zhang J, Crawford YG, Gauthier ML, Fordyce CA, McDermott KM, et al. Genetic and epigenetic changes in mammary epithelial cells identify a subpopulation of cells involved in early carcinogenesis. Cold Spring Harb Symp Quant Biol 2005; 70:317-27.
- 29. Dalla-Favera R, Gelmann EP, Gallo RC, Wong-Staal F. A human onc gene homologous to the transforming gene (v-sis) of simian sarcoma virus. Nature 1981; 292:31-5.
- Langbeheim H, Shih TY, Scolnick EM. Identification of a normal vertebrate cell protein related to the p21 src of Harvey murine sarcoma virus. Virology 1980; 106:292-300.
- Hahn WC, Counter CM, Lundberg AS, Beijersbergen RL, Brooks MW, Weinberg RA. Creation of human tumour cells with defined genetic elements. Nature 1999; 400:464-8.
- Stewart TA, Pattengale PK, Leder P. Spontaneous mammary adenocarcinomas in transgenic mice that carry and express MTV/myc fusion genes. Cell 1984; 38:627-37.

- Chin L, Tam A, Pomerantz J, Wong M, Holash J, Bardeesy N, et al. Essential role for oncogenic Ras in tumour maintenance. Nature 1999; 400:468-72.
- Crook T, Morgenstern JP, Crawford L, Banks L. Continued expression of HPV-16 E7 protein is required for maintenance of the transformed phenotype of cells co-transformed by HPV-16 plus EJ-ras. EMBO J 1989; 8:513-9.
- 35. Hanahan D, Weinberg RA. The hallmarks of cancer. Cell 2000; 100:57-70.
- 36. Fidler IJ. Selection of successive tumour lines for metastasis. Nat New Biol 1973; 242:148-9.
- Hanahan D. Dissecting multistep tumorigenesis in transgenic mice. Annu Rev Genet 1988; 22:479-519.
- 38. Nowell PC. The clonal evolution of tumor cell populations. Science 1976; 194:23-8.
- Beerenwinkel N, Antal T, Dingli D, Traulsen A, Kinzler KW, Velculescu VE, et al. Genetic progression and the waiting time to cancer. PLoS Comput Biol 2007; 3:225.
- Jones S, Chen WD, Parmigiani G, Diehl F, Beerenwinkel N, Antal T, et al. Comparative lesion sequencing provides insights into tumor evolution. Proc Natl Acad Sci USA 2008; 105:4283-8.
- Wood LD, Parsons DW, Jones S, Lin J, Sjoblom T, Leary RJ, et al. The genomic landscapes of human breast and colorectal cancers. Science 2007; 318:1108-13.
- Lininger RA, Fujii H, Man YG, Gabrielson E, Tavassoli FA. Comparison of loss of heterozygosity in primary and recurrent ductal carcinoma in situ of the breast. Mod Pathol 1998; 11:1151-9.
- Vos CBJ, Cleton-Jansen AM, Berx G, de Leeuw WJF, ter Haar NT, van Roy F, et al. E-cadherin inactivation in lobular carcinoma in situ of the breast: an early event in tumorigenesis. Br J Cancer 1997; 76:1131-3.
- 44. Bettendorf O, Schmidt H, Staebler A, Grobholz R, Heinecke A, Boecker W, et al. Chromosomal imbalances, loss of heterozygosity and immunohistochemical expression of TP53, RB1 and PTEN in intraductal cancer, intraepithelial neoplasia and invasive adenocarcinoma of the prostate. Genes Chromosomes Cancer 2008; 47:565-72.
- 45. Larson PS, de las Morenas A, Cerda SR, Bennett SR, Cupples LA, Rosenberg CL. Quantitative analysis of allele imbalance supports atypical ductal hyperplasia lesions as direct breast cancer precursors. J Pathol 2006; 209:307-16.
- Soto AM, Vandenberg LN, Maffini MV, Sonnenschein C. Does breast cancer start in the womb? Basic Clin Pharmacol Toxicol 2008; 102:125-33.
- Soto AM, Maffini MV, Sonnenschein C. Neoplasia as development gone awry: the role of endocrine disruptors. Int J Androl 2008; 31:288-93.
- 48. Virchow RL. Rudolph Virchow on ochronosis, 1866. Arthritis Rheum 1966; 9:66-71.
- 49. Balkwill F, Mantovani A. Inflammation and cancer: back to Virchow? Lancet 2001; 357:539-45.
- 50. Karin M, Lawrence T, Nizet V. Innate immunity gone awry: linking microbial infections to chronic inflammation and cancer. Cell 2006; 124:823-35.
- Lin WW, Karin M. A cytokine-mediated link between innate immunity, inflammation and cancer. J Clin Invest 2007; 117:1175-83.
- Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. Nature 2008; 454:436-44.
- Paget S. The distributions of secondary growths in cancer of the breast. Lancet 1889; 1:571-3.
- Fidler IJ, Yano S, Zhang RD, Fujimaki T, Bucana CD. The seed and soil hypothesis: vascularisation and brain metastases. Lancet Oncol 2002; 3:53-7.
- Billingham RE, Orr JW, Woodhouse DL. Transplantation of skin components during chemical carcinogenesis with 20-methylcholanthrene. Br J Cancer 1951; 5:417-32.
- Dolberg DS, Hollingsworth R, Hertle M, Bissell MJ. Wounding and its role in RSVmediated tumor formation. Science 1985; 230:676-8.
- 57. Dolberg DS, Bissell MJ. Inability of Rous sarcoma virus to cause sarcomas in the avian embryo. Nature 1984; 309:552-6.
- Ames BN, Gurney EG, Miller JA, Bartsch H. Carcinogens as frameshift mutagens: metabolites and derivatives of 2-acetylaminofluorene and other aromatic amine carcinogens. Proc Natl Acad Sci USA 1972; 69:3128-32.
- 59. Niehrs C. Axis formation: redundancy rules. Curr Biol 2005; 15:391-3.
- Niehrs C. Regionally specific induction by the Spemann-Mangold organizer. Nat Rev Genet 2004; 5:425-34.
- Beetschen JC, Duprat AM. Contrasting influences of the organizer and induction concepts on the scientific activity of French embryologists. Int J Dev Biol 2001; 45:73-81.
- Dvorak HF. Tumors: wounds that do not heal. Similarities between tumor stroma generation and wound healing. N Engl J Med 1986; 315:1650-9.
- Buess M, Nuyten DS, Hastie T, Nielsen T, Pesich R, Brown PO. Characterization of heterotypic interaction effects in vitro to deconvolute global gene expression profiles in cancer. Genome Biol 2007; 8:191.
- 64. Fromigue O, Louis K, Dayem M, Milanini J, Pages G, Tartare-Deckert S, et al. Gene expression profiling of normal human pulmonary fibroblasts following coculture with nonsmall-cell lung cancer cells reveals alterations related to matrix degradation, angiogenesis, cell growth and survival. Oncogene 2003; 22:8487-97.
- Gallagher PG, Bao Y, Prorock A, Zigrino P, Nischt R, Politi V, et al. Gene expression profiling reveals cross-talk between melanoma and fibroblasts: implications for host-tumor interactions in metastasis. Cancer Res 2005; 65:4134-46.
- Sato N, Maehara N, Goggins M. Gene expression profiling of tumor-stromal interactions between pancreatic cancer cells and stromal fibroblasts. Cancer Res 2004; 64:6950-6.

- 67. Lacina L, Dvorankova B, Smetana K Jr, Chovanec M, Plzak J, Tachezy R, et al. Marker profiling of normal keratinocytes identifies the stroma from squamous cell carcinoma of the oral cavity as a modulatory microenvironment in co-culture. Int J Radiat Biol 2007; 83:837-48.
- Miki Y, Suzuki T, Tazawa C, Yamaguchi Y, Kitada K, Honma S, et al. Aromatase localization in human breast cancer tissues: possible interactions between intratumoral stromal and parenchymal cells. Cancer Res 2007; 67:3945-54.
- Mahadevan D, Von Hoff DD. Tumor-stroma interactions in pancreatic ductal adenocarcinoma. Mol Cancer Ther 2007; 6:1186-97.
- Wang J, Levenson AS, Satcher RL Jr. Identification of a unique set of genes altered during cell-cell contact in an in vitro model of prostate cancer bone metastasis. Int J Mol Med 2006; 17:849-56.
- Bavik C, Coleman I, Dean JP, Knudsen B, Plymate S, Nelson PS. The gene expression program of prostate fibroblast senescence modulates neoplastic epithelial cell proliferation through paracrine mechanisms. Cancer Res 2006; 66:794-802.
- 72. Rich JN, Hans C, Jones B, Iversen ES, McLendon RE, Rasheed BK, et al. Gene expression profiling and genetic markers in glioblastoma survival. Cancer Res 2005; 65:4051-8.
- 73. Spentzos D, Levine DA, Ramoni MF, Joseph M, Gu X, Boyd J, et al. A gene expression signature with independent prognostic significance in epithelial ovarian cancer. J Clin Oncol 2004; 22:4700-10.
- Tothill RW, Tinker AV, George J, Brown R, Fox SB, Lade S, et al. Novel molecular subtypes of serous and endometrioid ovarian cancer linked to clinical outcome. Clin Cancer Res 2008; 14:5198-208.
- 75. Sommer SS. Does cancer kill the individual and save the species? Hum Mutat 1994; 3:166-9.
- Pollard JW. Macrophages define the invasive microenvironment in breast cancer. J Leukoc Biol 2008; 84:623-30.
- Radisky DC, Levy DD, Littlepage LE, Liu H, Nelson CM, Fata JE, et al. Rac1b and reactive oxygen species mediate MMP-3-induced EMT and genomic instability. Nature 2005; 436:123-7.
- Pikarsky E, Porat RM, Stein I, Abramovitch R, Amit S, Kasem S, et al. NF[kappa]B functions as a tumour promoter in inflammation-associated cancer. Nature 2004; 431:461-6.
- Bean GR, Bryson AD, Pilie PG, Goldenberg V, Baker JC Jr, Ibarra C, et al. Morphologically normal-appearing mammary epithelial cells obtained from high-risk women exhibit methylation silencing of INK4a/ARF. Clin Cancer Res 2007; 13:6834-41.
- Kuperwasser C, Chavarria T, Wu M, Magrane G, Gray JW, Carey L, et al. Reconstruction of functionally normal and malignant human breast tissues in mice. Proc Natl Acad Sci USA 2004; 101:4966-71.
- Maffini MV, Soto AM, Calabro JM, Ucci AA, Sonnenschein C. The stroma as a crucial target in rat mammary gland carcinogenesis. J Cell Sci 2004; 117:1495-502.
- 82. Campisi J. Fragile fugue: p53 in aging, cancer and IGF signaling. Nat Med 2004; 10:231-2.
- Campisi J. Aging, tumor suppression and cancer: high wire-act! Mech Ageing Dev 2005; 126:51-8.
- Krtolica A, Campisi J. Cancer and aging: a model for the cancer promoting effects of the aging stroma. Int J Biochem Cell Biol 2002; 34:1401-14.
- Parrinello S, Coppe JP, Krtolica A, Campisi J. Stromal-epithelial interactions in aging and cancer: senescent fibroblasts alter epithelial cell differentiation. J Cell Sci 2005; 118:485-96.
- Ellis MJ, Singer C, Hornby A, Rasmussen A, Cullen KJ. Insulin-like growth factor mediated stromal-epithelial interactions in human breast cancer. Breast Cancer Res Treat 1994; 31:249-61.
- Matsumoto K, Nakamura T. Hepatocyte growth factor and the Met system as a mediator of tumor-stromal interactions. Int J Cancer 2006; 119:477-83.
- Ostman A. PDGF receptors-mediators of autocrine tumor growth and regulators of tumor vasculature and stroma. Cytokine Growth Factor Rev 2004; 15:275-86.
- Wisniewski HG, Hua JC, Poppers DM, Naime D, Vilcek J, Cronstein BN. TNF/IL-1inducible protein TSG-6 potentiates plasmin inhibition by inter-alpha-inhibitor and exerts a strong anti-inflammatory effect in vivo. J Immunol 1996; 156:1609-15.
- Joesting MS, Perrin S, Elenbaas B, Fawell SE, Rubin JS, Franco OE, et al. Identification of SFRP1 as a candidate mediator of stromal-to-epithelial signaling in prostate cancer. Cancer Res 2005; 65:10423-30.
- Moinfar F, Man YG, Arnould L, Bratthauer GL, Ratschek M, Tavassoli FA. Concurrent and independent genetic alterations in the stromal and epithelial cells of mammary carcinoma: implications for tumorigenesis. Cancer Res 2000; 60:2562-6.
- Fukino K, Shen L, Matsumoto S, Morrison CD, Mutter GL, Eng C. Combined total genome loss of heterozygosity scan of breast cancer stroma and epithelium reveals multiplicity of stromal targets. Cancer Res 2004; 64:7231-6.
- Kurose K, Gilley K, Matsumoto S, Watson PH, Zhou XP, Eng C. Frequent somatic mutations in PTEN and TP53 are mutually exclusive in the stroma of breast carcinomas. Nat Genet 2002; 32:355-7.
- Plon SE, Pirics ML, Nuchtern J, Hicks J, Russell H, Agrawal S, et al. Multiple tumors in a child with germ-line mutations in TP53 and PTEN. N Engl J Med 2008; 359:537-9.
- Patocs A, Zhang L, Xu Y, Weber F, Caldes T, Mutter GL, et al. Breast-cancer stromal cells with TP53 mutations and nodal metastases. N Engl J Med 2007; 357:2543-51.
- Studeny M, Marini FC, Dembinski JL, Zompetta C, Cabreira-Hansen M, Bekele BN, et al. Mesenchymal stem cells: potential precursors for tumor stroma and targeted-delivery vehicles for anticancer agents. J Natl Cancer Inst 2004; 96:1593-603.
- Karnoub AE, Dash AB, Vo AP, Sullivan A, Brooks MW, Bell GW, et al. Mesenchymal stem cells within tumour stroma promote breast cancer metastasis. Nature 2007; 449:557-63.

- 98. Weinberg RA. Coevolution in the tumor microenvironment. Nat Genet 2008; 40:494-5.
- Littlepage LE, Egeblad M, Werb Z. Coevolution of cancer and stromal cellular responses. Cancer Cell 2005; 7:499-500.
- 100. Campisi J. Suppressing cancer: the importance of being senescent. Science 2005; 309:886-7.
- Sherr CJ. An Arf(GFP/GFP) reporter mouse reveals that the Arf tumor suppressor monitors latent oncogenic signals in vivo. Cell Cycle 2004; 3:239-40.
- Palmero I, Serrano M. Induction of senescence by oncogenic Ras. Methods Enzymol 2001; 333:247-56.
- Brenner AJ, Stampfer MR, Aldaz CM. Increased p16 expression with first senescence arrest in human mammary epithelial cells and extended growth capacity with p16 inactivation. Oncogene 1998; 17:199-205.
- Rose DW, McCabe G, Feramisco JR, Adler M. Expression of c-fos and AP-1 activity in senescent human fibroblasts is not sufficient for DNA synthesis. J Cell Biol 1992; 119:1405-11.
- Yu GL, Bradley JD, Attardi LD, Blackburn EH. In vivo alteration of telomere sequences and senescence caused by mutated Tetrahymena telomerase RNAs. Nature 1990; 344:126-32.
- Blagosklonny MV, Campisi J. Cancer and aging: more puzzles, more promises? Cell Cycle 2008; 7:2615-8.
- 107. Campisi J. Aging and cancer cell biology, 2008. Aging Cell 2008; 7:281-4.
- Campisi J, d'Adda di Fagagna F. Cellular senescence: when bad things happen to good cells. Nat Rev Mol Cell Biol 2007; 8:729-40.
- Knudson AG Jr. Mutation and cancer: statistical study of retinoblastoma. Proc Natl Acad Sci USA 1971; 68:820-3.
- 110. Vijg J. Somatic mutations and aging: a re-evaluation. Mutat Res 2000; 447:117-35.
- 111. Mendelsohn ML. Somatic cell mutation as a radiation biodosimeter and predictor of cancer risk and aging. Jpn J Cancer Res 1996; 87.
- 112. Alexander P. The relationship between ageing and cancer: somatic mutations or break-down of host defence mechanisms. Bull Schweiz Akad Med Wiss 1969; 24:258-71.
- 113. Qu T, Walston JD, Yang H, Fedarko NS, Xue QL, Beamer BA, et al. Upregulated ex vivo expression of stress-responsive inflammatory pathway genes by LPS-challenged CD14(+) monocytes in frail older adults. Mech Ageing Dev 2008.
- 114. Campisi J. Aging and cancer: the double-edged sword of replicative senescence. J Am Geriatr Soc 1997; 45:482-8.
- 115. Perez-Pinera P, Chang Y, Deuel TF. Pleiotrophin, a multifunctional tumor promoter through induction of tumor angiogenesis, remodeling of the tumor microenvironment, and activation of stromal fibroblasts. Cell Cycle 2007; 6:2877-83.
- 116. Parrott JA, Kim G, Mosher R, Skinner MK. Expression and action of keratinocyte growth factor (KGF) in normal ovarian surface epithelium and ovarian cancer. Mol Cell Endocrinol 2000; 167:77-87.
- 117. Parrott JA, Kim G, Skinner MK. Expression and action of kit ligand/stem cell factor in normal human and bovine ovarian surface epithelium and ovarian cancer. Biol Reprod 2000; 62:1600-9.
- Parrott JA, Nilsson E, Mosher R, Magrane G, Albertson D, Pinkel D, et al. Stromal-epithelial interactions in the progression of ovarian cancer: influence and source of tumor stromal cells. Mol Cell Endocrinol 2001; 175:29-39.
- Choi KC, Kang SK, Tai CJ, Auersperg N, Leung PC. The regulation of apoptosis by activin and transforming growth factor-beta in early neoplastic and tumorigenic ovarian surface epithelium. J Clin Endocrinol Metab 2001; 86:2125-35.
- Hu YL, Tee MK, Goetzl EJ, Auersperg N, Mills GB, Ferrara N, et al. Lysophosphatidic acid induction of vascular endothelial growth factor expression in human ovarian cancer cells. J Natl Cancer Inst 2001; 93:762-8.
- Choi KC, Kang SK, Tai CJ, Auersperg N, Leung PC. Follicle-stimulating hormone activates mitogen-activated protein kinase in preneoplastic and neoplastic ovarian surface epithelial cells. J Clin Endocrinol Metab 2002; 87:2245-53.
- 122. Zand L, Qiang F, Roskelley CD, Leung PC, Auersperg N. Differential effects of cellular fibronectin and plasma fibronectin on ovarian cancer cell adhesion, migration and invasion. In Vitro Cell Dev Biol Anim 2003; 39:178-82.
- Kaminski A, Hahne JC, Haddouti el M, Florin A, Wellmann A, Wernert N. Tumourstroma interactions between metastatic prostate cancer cells and fibroblasts. Int J Mol Med 2006; 18:941-50.
- 124. Kataoka H, Tanaka H, Nagaike K, Uchiyama S, Itoh H. Role of cancer cell-stroma interaction in invasive growth of cancer cells. Hum Cell 2003; 16:1-14.
- 125. Tang Y, Nakada MT, Kesavan P, McCabe F, Millar H, Rafferty P, et al. Extracellular matrix metalloproteinase inducer stimulates tumor angiogenesis by elevating vascular endothelial cell growth factor and matrix metalloproteinases. Cancer Res 2005; 65:3193-9.
- 126. Acuff HB, Carter KJ, Fingleton B, Gorden DL, Matrisian LM. Matrix metalloproteinase-9 from bone marrow-derived cells contributes to survival but not growth of tumor cells in the lung microenvironment. Cancer Res 2006; 66:259-66.
- Bruner KL, Rodgers WH, Gold LI, Korc M, Hargrove JT, Matrisian LM, et al. Transforming growth factor beta mediates the progesterone suppression of an epithelial metalloproteinase by adjacent stroma in the human endometrium. Proc Natl Acad Sci USA 1995; 92:7362-6.
- 128. Koukourakis MI, Giatromanolaki A, Brekken RA, Sivridis E, Gatter KC, Harris AL, et al. Enhanced expression of SPARC/osteonectin in the tumor-associated stroma of non-small cell lung cancer is correlated with markers of hypoxia/acidity and with poor prognosis of patients. Cancer Res 2003; 63:5376-80.

- 129. Hu M, Yao J, Carroll DK, Weremowicz S, Chen H, Carrasco D, et al. Regulation of in situ to invasive breast carcinoma transition. Cancer Cell 2008; 13:394-406.
- Pietras K, Pahler J, Bergers G, Hanahan D. Functions of paracrine PDGF signaling in the proangiogenic tumor stroma revealed by pharmacological targeting. PLoS Med 2008; 5:19.
- 131. Fearon ER, Vogelstein BA. A genetic model for colorectal tumorigenesis. Cell 1990; 61:759-67.
- 132. Tuhkanen H, Anttila M, Kosma VM, Yla-Herttuala S, Heinonen S, Kuronen A, et al. Genetic alterations in the peritumoral stromal cells of malignant and borderline epithelial ovarian tumors as indicated by allelic imbalance on chromosome 3p. Int J Cancer 2004; 109:247-52.
- 133. Blagosklonny MV. Molecular theory of cancer. Cancer Biol Ther 2005; 4:621-7.
- Allinen M, Beroukhim R, Cai L, Brennan C, Lahti-Domenici J, Huang H, et al. Molecular characterization of the tumor microenvironment in breast cancer. Cancer Cell 2004; 6:17-32.
- 135. Tanaka H, Masuda T, Tokuoka S, Takahashi Y, Komai M, Nagao K, et al. Time course study on the development of allergen-induced airway remodeling in mice: the effect of allergen avoidance on established airway remodeling. Inflamm Res 2002; 51:307-16.
- 136. Yang WL, Godwin AK, Xu XX. Tumor necrosis factor-alpha-induced matrix proteolytic enzyme production and basement membrane remodeling by human ovarian surface epithelial cells: molecular basis linking ovulation and cancer risk. Cancer Res 2004; 64:1534-40.
- 137. Lama VN, Phan SH. The extrapulmonary origin of fibroblasts: stem/progenitor cells and beyond. Proc Am Thorac Soc 2006; 3:373-6.
- 138. Sangai T, Ishii G, Kodama K, Miyamoto S, Aoyagi Y, Ito T, et al. Effect of differences in cancer cells and tumor growth sites on recruiting bone marrow-derived endothelial cells and myofibroblasts in cancer-induced stroma. Int J Cancer 2005; 115:885-92.
- Gao D, Nolan DJ, Mellick AS, Bambino K, McDonnell K, Mittal V. Endothelial progenitor cells control the angiogenic switch in mouse lung metastasis. Science 2008; 319:195-8.
- 140. Sugimoto H, Mundel TM, Kieran MW, Kalluri R. Identification of fibroblast heterogeneity in the tumor microenvironment. Cancer Biol Ther 2006; 5:1640-6.